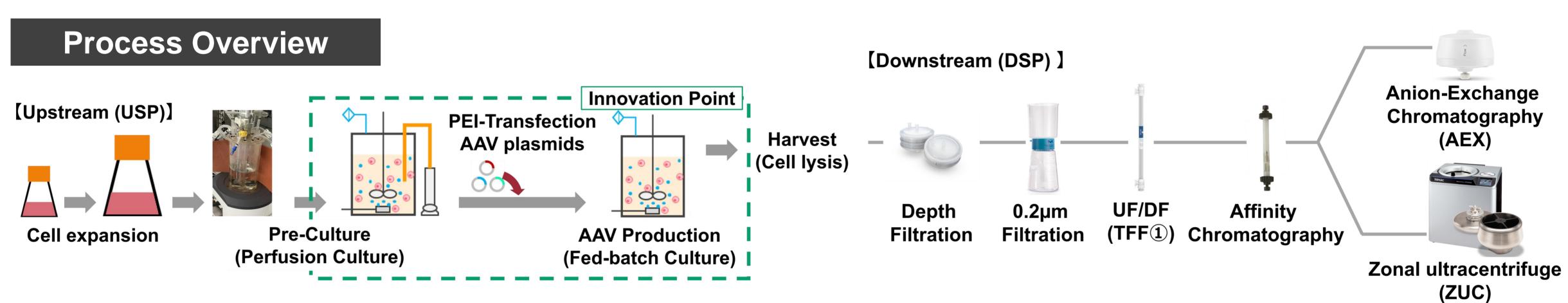
# Partners for Life

# Cost-effective rAAV mass manufacturing by high-density cell with small-scale equipment

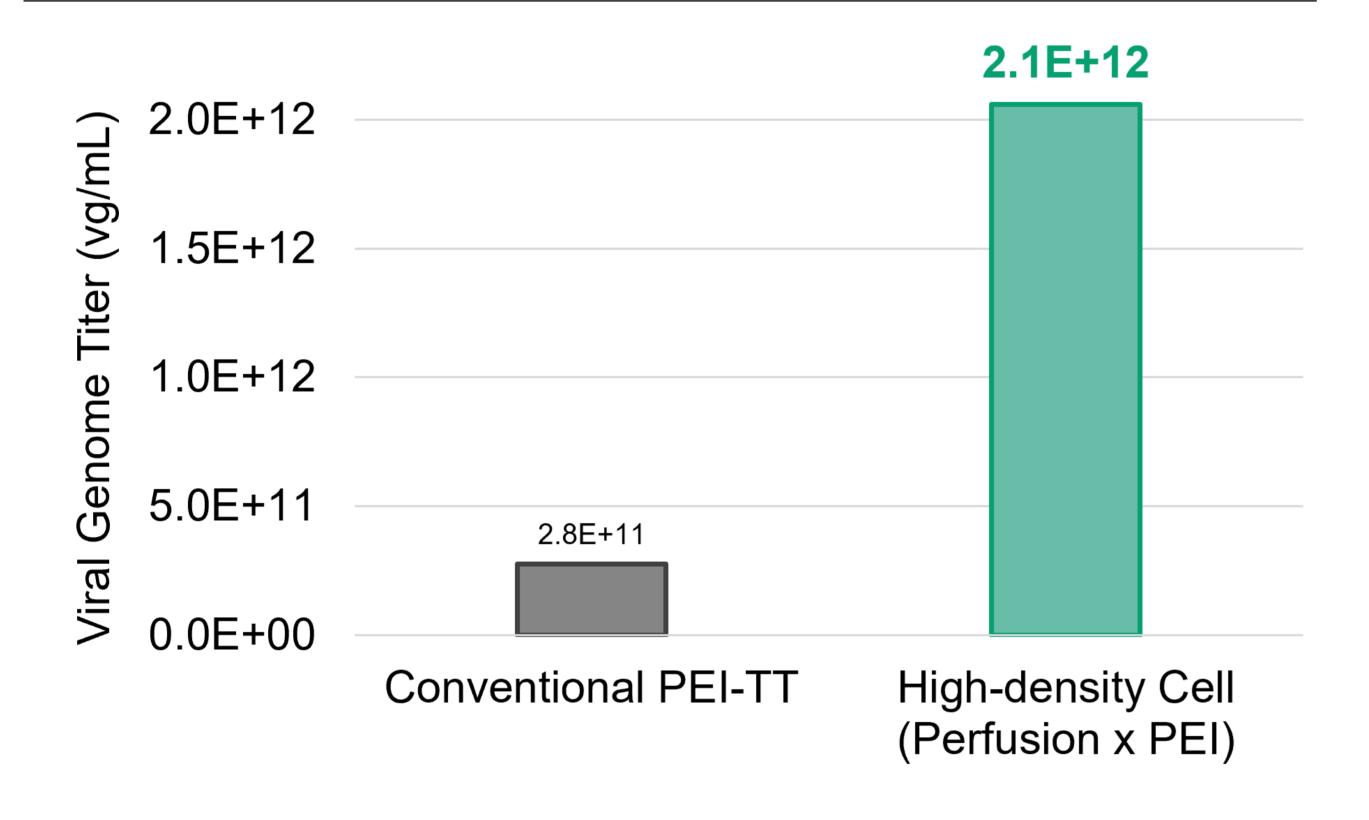
Conventional PEI-TT methods (PEI-Transfection to 2E6 cell/mL) have been considered low Background in rAAV productivity due to the limitation of cell density. As a result, scale-up production in a large-scale equipment is required for AAV mass production.

To increase productivity, our intensified AAV production method enables PEI-mediated Highlights transfection into high-density cells by increasing cell density through perfusion culture. It offers approx.10-fold higher rAAV5 productivity in a single transfection batch. This enables us to provide clients with high-yield AAV titers using small-scale equipment, which are equivalent to results obtained with large-scale equipment.

We also demonstrated several downstream purification processes using samples derived from high-density cells. Even with high-density cells, hcDNA and HCP were reduced to the low-density level as those obtained by conventional PEI-TT methods. By replacing the Full/Empty separation with Zonal Ultracentrifugation (ZUC) method, the full ration is significantly improved.

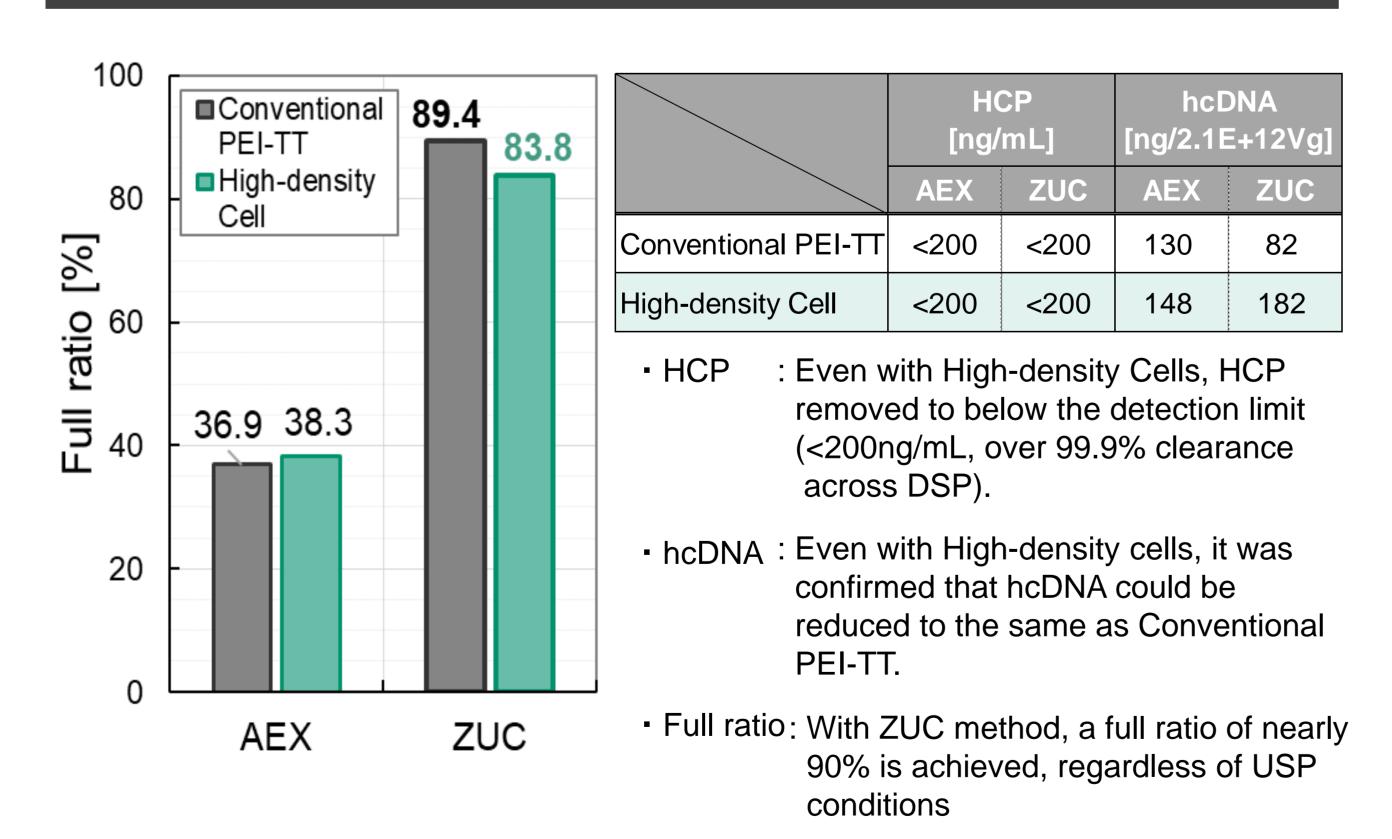


### Intensified rAAV5 production from high-density cell



#### AAV production Pre-culture PEI-TF (Fed-batch) (Perfusion) 40.0 100 35.0 95 30.0 90 Viability VCD 25.0 85 20.0 15.0 Perfusion VCD Start 70 10.0 5.0 65 0.0 (culture day) 8 AAV5 genome titer Productivity fold change Method (cell/mL) Vg/mL base Vg base (Vg/mL) Conventional PEI-TT 2.0E+06 2.8E+11 **High-density Cell** 9.9 2.1E+12 2.1E+07

# Impurities clearance and full particle ratio at ZUC



<sup>\*</sup> Above is our lab scale data. Full/Empty separation by AEX can be further improved depending on the yield.

# Potential reduction amount of plasmid

7.5

		Scale	Titer	Total VG @USP	Plasmid amount
		(L)	(Vg/L)	(Vg)	(mg per 2.8E16vg)
	Conventional-TT	100	2.8E+14	2.8E+16	100
	HCD-PEI	10	2.1E+15	2.1E+16	49 51%

This method is estimated to reduce the plasmid by 51% to obtain the same amount of rAAV produced in a 100L batch at a 10L scale without any equipment and steps other than perfusion culture.

#### **Further improvement** Single perfusion culture for Repeat 10-fold batches cell supply in mother reactor in daughter reactors TF Day1 TF Day2 50% volume Cell bleeding and Transfection TF Day3 supplementing culture medium

This method enabling 10-fold rAAV production with high-density cells can be repeatedly implemented by preparing another reactors. It could potentially provide productivity equivalent to equipment over 100 -fold larger, even with a small-scale reactor.





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(Perfusion x PEI)

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